

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/232008176>

Llácer, E., Santiago-Álvarez, C., and Jacas, J. A. Could sterile males be used to vector a microbiological control agent? The case of *Rhynchophorus ferrugineus* and *Beauveria bassia*...

ARTICLE in BULLETIN OF ENTOMOLOGICAL RESEARCH · OCTOBER 2012

Impact Factor: 1.91 · DOI: 10.1017/S0007485312000582 · Source: PubMed

CITATIONS

11

READS

57

3 AUTHORS, INCLUDING:



Elena Llácer

Valencian Institute for Agricultural Resea...

28 PUBLICATIONS 347 CITATIONS

SEE PROFILE



J. A. Jacas

Universitat Jaume I

204 PUBLICATIONS 1,641 CITATIONS

SEE PROFILE

Bulletin of Entomological Research

<http://journals.cambridge.org/BER>

Additional services for ***Bulletin of Entomological Research***:

Email alerts: [Click here](#)

Subscriptions: [Click here](#)

Commercial reprints: [Click here](#)

Terms of use : [Click here](#)



Could sterile males be used to vector a microbiological control agent? The case of *Rhynchophorus ferrugineus* and *Beauveria bassiana*

E. Llácer, C. Santiago-Álvarez and J.A. Jacas

Bulletin of Entomological Research / *FirstView* Article / October 2012, pp 1 - 10

DOI: 10.1017/S0007485312000582, Published online:

Link to this article: http://journals.cambridge.org/abstract_S0007485312000582

How to cite this article:

E. Llácer, C. Santiago-Álvarez and J.A. Jacas Could sterile males be used to vector a microbiological control agent? The case of *Rhynchophorus ferrugineus* and *Beauveria bassiana*. Bulletin of Entomological Research, Available on CJO doi:10.1017/S0007485312000582

Request Permissions : [Click here](#)

Could sterile males be used to vector a microbiological control agent? The case of *Rhynchophorus ferrugineus* and *Beauveria bassiana*

E. Llácer^{1*}, C. Santiago-Álvarez² and J.A. Jacas³

¹Instituto Valenciano de Investigaciones Agrarias (IVIA), Unidad Asociada de Entomología Agrícola UJI-IVIA, Centro de Protección Vegetal y Biotecnología, Carretera Montcada-Nàquera km 4.5, E-46113-Montcada, Spain: ²Universidad de Córdoba, Departamento de Ciencias y Recursos Agrícolas y Forestales, ETSIAM, E-14071-Córdoba, Spain: ³Universitat Jaume I (UJI), Unitat Associada d'Entomologia Agrícola UJI-IVIA, Departament de Ciències Agràries i del Medi Natural, Campus del Riu Sec, Avenida Vicent Sos Baynat s/n, E-12071-Castelló de la Plana, Spain

Abstract

Rhynchophorus ferrugineus (Coleoptera, Curculionidae) is the most threatening pest of palms worldwide. The potential of gamma-irradiated males to spread a pathogenic strain of the entomopathogenic fungus *Beauveria bassiana* (Ascomycota: Clavicipitaceae) to control this pest was studied. First, the effects of gamma irradiation (15 and 25 Gy) on the mating success and performance of adult males irradiated at age one day were studied in the laboratory. Although male longevity decreased after irradiation (118.6 vs. 244.7 days for irradiated and control males, respectively) and their testes suffered from the treatment, fecundity of mated females did not depend on the irradiation status of the male (86.8 ± 5.5 eggs in 15 days). However, egg hatching was significantly lower in couples with irradiated males (31.4% vs. 86.5% for irradiated and control couples, respectively), and this value decreased after a second mating (6.1% vs. 85.9%). Therefore, irradiation did not affect male sexual competitiveness but sperm quality. Second, a semi-field assay was carried out to evaluate infestation in young *Phoenix canariensis* caused by different combinations of couples with irradiated and/or *B. bassiana*-challenged males. The number of immature stages found in infested palms was significantly higher when females mated with untreated males and lower when mated with irradiated males (either *B. bassiana*-infected or not). Some females from the fungus-challenged treatments showed post-mortem hyphal growth, and this horizontal transmission proves that irradiated males could act as a vector for *B. bassiana* and should be considered as a new method to improve the biological control of *R. ferrugineus*.

Keywords: gamma irradiation, horizontal transmission, entomopathogenic fungus, palm pest, *Phoenix canariensis*, autodissemination, sterile males

(Accepted 13 July 2012)

*Author for correspondence
Fax: 00 34 96 3424001
E-mail: ellacer@hotmail.com

Introduction

Nowadays, *Rhynchophorus ferrugineus* (Olivier) (Coleoptera, Curculionidae) is considered the main pest of palms in the Middle East and the Mediterranean Basin (Faleiro, 2006; Ll  cer & Jacas, 2010), and it may become a serious pest in America (European and Mediterranean Plant Protection Organization, 2009). This species is a concealed tissue borer that lives inside the palm. Adults often remain and extensively reproduce within the same host until the larvae have destroyed the growing tip of the palm, causing the palm to die. *Rhynchophorus ferrugineus* feeds on soft tissues of most Palmaceae, and the Canary Islands Date Palm, *Phoenix canariensis* Hort. ex Chabaud, a species widely used as ornamental, is its preferred host in the Mediterranean Basin (Dembilio *et al.*, 2009). One of the main handicaps for the efficient control of *R. ferrugineus* is its cryptic habits. Current control methods are mainly based on the application of large quantities of insecticides designed to limit the infestation in treated palms. These procedures have been improved during the latest decade to enhance efficacy and to reduce application costs and toxicity to non-target organisms (Faleiro, 2006; Ll  cer *et al.*, 2009, 2012; Dembilio *et al.*, 2010a,b). Mass trapping is also widely used against this pest (Abbas *et al.*, 2006; Faleiro & Kumar, 2008). Another useful procedure for pest control, the Sterile Insect Technique (Lance & McInnis, 2005), has been tested against *R. ferrugineus*. Chemosterilization (Rahalkar *et al.*, 1975), gamma (Ramachandran, 1991) and X-ray radiation (Rahalkar *et al.*, 1973) proved successful in the laboratory to reduce egg hatching when sterilized males were allowed to mate with untreated females. Field studies were also carried out with promising results (Ranavara *et al.*, 1975; Rahalkar *et al.*, 1977). More recently, gamma-sterilized males have been used in different studies on the basic biology of *R. ferrugineus* (Gothi *et al.*, 2007; Al-Ayedh & Rassol, 2009; Prabhu *et al.*, 2010). These studies point that gamma-irradiation of males induces permanent sterility and has no influence on mating behaviour, one of the requisites for the successful application of the SIT (Lance & McInnis, 2005). Even if the polygamy of *R. ferrugineus* females and the extremely high cost of this technique makes the SIT hardly sustainable at this moment, gamma-irradiation of males has features that could make it a useful tool to control this pest.

Although few studies have been conducted on the natural enemies of *R. ferrugineus* or other *Rhynchophorus* species (Murphy & Briscoe, 1999; Faleiro, 2006), biological control is another interesting strategy against this pest. Entomopathogenic nematodes (EPNs) are safe for non-target vertebrates and for the environment and have proved effective against *R. ferrugineus* (Abbas *et al.*, 2001a,b; Saleh & Alheji, 2003; Elawad *et al.*, 2007; Ll  cer *et al.*, 2009; Dembilio *et al.*, 2010a). In addition to EPNs, entomopathogenic fungi (EPF) can also provide an excellent alternative to chemical control. EPFs infect the host by direct contact and by horizontal transmission from infected insects or cadavers to healthy insects (Lacey *et al.*, 1999; Quesada-Moraga *et al.*, 2004). These unique characters make EPFs especially important for the control of concealed insects as *R. ferrugineus*. Different strains of *Metarhiziumanisopliae* (Metschnikoff) Sokorin (Ascomycota: Clavicipitaceae) and *Beauveria bassiana* (Balsamo) Vuillemin (Ascomycota: Clavicipitaceae) have been tested against *R. ferrugineus* (Gindin *et al.*, 2006; El-Sufty *et al.*, 2009; Sewify *et al.*, 2009; Dembilio *et al.*, 2010b). Recently, El-Sufty *et al.* (2009) successfully reduced the incidence of *R. ferrugineus*

under field conditions in Egypt using an indigenous isolate of *B. bassiana*, as did Dembilio *et al.* (2010b) in Spain under semi-field conditions (efficacies around 85%). The latter authors demonstrated for the *B. bassiana* strain CECT-20752 that fungus-challenged males were able to transmit the disease to healthy females in the laboratory and this infection resulted in a subsequent reduction in fecundity and egg hatching. At the moment, it is necessary to find methods to apply *B. bassiana* in field so that it results effective in controlling the pest. Therefore, we decided to investigate the potential of using gamma-irradiated males to spread the disease, as suggested for other pests of economic importance (Vickers *et al.*, 2004; Novelo-Rinc  n *et al.*, 2009). To achieve this goal, we have studied the reproductive biology of *R. ferrugineus* adult females obtained from *P. canariensis*-fed immature stages and specifically the possible occurrence of parthenogenesis, the effects of gamma irradiation on the mating success and performance of *R. ferrugineus* males. Further, we have studied the efficacy of gamma-sterilized males to disperse *B. bassiana* under controlled conditions. These studies should pave the way for developing sound biological control strategies against *R. ferrugineus* based on the use of sterile males to vector EPFs.

Materials and methods

The assays reported in this study were carried out at the Instituto Valenciano de Investigaciones Agrarias (IVIA, Montcada, Spain). Laboratory trials took place in a climatic cabinet at $25 \pm 2^\circ\text{C}$ and $80 \pm 10\%$ RH and a photoperiod of 16:8 L:D. Semi-field trials were performed in a double mesh security enclosure protected from the rain by a plastic roof and containing 24 independent cages ($4 \times 3 \times 3\text{m}$) under natural light and temperature conditions during August (mean temperature $25.0 \pm 0.3^\circ\text{C}$; max, 28.8°C ; min., 21.9°C) and October (mean temperature $15.4 \pm 0.4^\circ\text{C}$; max, 21.4°C ; min., 12.2°C) 2010.

Experimental insects

Adult weevils used in this study were obtained from cocoons collected in naturally infested *P. canariensis* palms around the city of Valencia, Spain. These cocoons were individually placed in 100-ml, plastic containers with perforated lids and incubated until adult emergence. Newly emerged adults were sexed and individually transferred to new containers as above. Filter paper and thin slices of red apple cultivar 'Starking delicious' were provided as food source and replaced weekly. Adults were maintained under these conditions (referred to as 'standard containers' henceforth) in a climatic cabinet until they were used in the assays. Unless otherwise stated, the same method and conditions apply to insects used in our laboratory experiments.

Gamma irradiation

One-day-old *R. ferrugineus* males were exposed in groups of six to a dose of 15 or 25 Gy of gamma rays using a Gammacell 220 SN 57R (Cobalt source) irradiator located in the nearby Medfly Mass Rearing Facility at Caudete de las Fuentes, Spain.

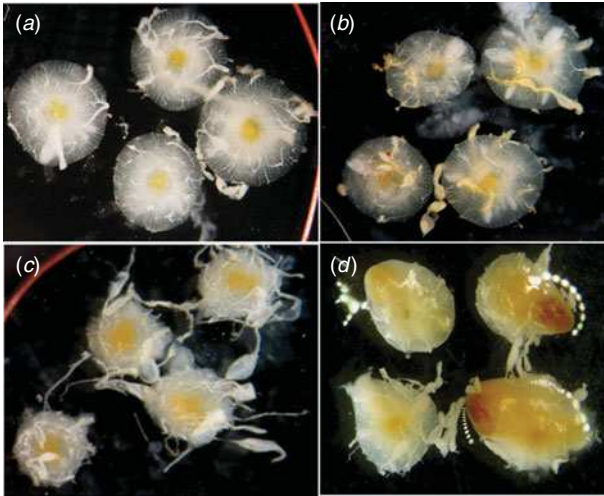


Fig. 1. Appearance of *R. ferrugineus* testes: (a) all two-bilobed testes are turgid and sparkling, (b) two lobes show lack of turgidity, (c) all two-bilobed testes are shrivelled, and (d) all two-bilobed testes show protuberances.

Laboratory trials

Female reproductive biology

Newly emerged females were individually introduced into a Petri dish (9 cm diameter and 2 cm height) either alone or with a five-day-old male and left undisturbed for 30 min. Afterwards, females were placed individually in 100-ml standard containers. Filter paper and thin slices of red apple cultivar 'Starking delicious' were provided as food source and oviposition substrate. Eggs were collected daily for 30 days using a fine paintbrush and transferred to a Petri dish (5 cm in diameter and 1.4 cm height) on artificial diet (Martin & Cabello, 2006) where egg hatching was scored five days later. Fifteen replicates (females) per treatment were considered and these results were compared by ANOVA.

Adult longevity

Longevities of untreated females, untreated males, and 15 and 25 Gy gamma-irradiated males were assessed. One-day-old adults (either irradiated or not) were individually placed in standard containers and maintained until they died. Two assays, including 17 replicates per treatment each, were considered. Longevity was analysed by ANOVA ($P < 0.05$) and, if necessary, means were separated using Duncan's test.

Mating success of irradiated males

Sixty one-day-old weevils per treatment (same treatments as for longevity) were individualized in standard containers. On a weekly basis for the first eight weeks, and then every four weeks until week 28, and every eight weeks from weeks 28 to 52, three adult males per treatment were selected (provided that there were survivors) and used in our experiments. Individualized males were allowed to mate with a 15-day-old female in a Petri dish (9 cm diameter and 2 cm height) for 30 min and the number of couples that were actually observed mating (apparent successful mating) was recorded.

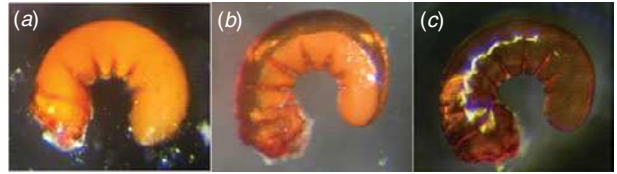


Fig. 2. Sperm mass in *R. ferrugineus* sclerotized spermatheca: (a) spermatheca nearly full of sperm, (b) spermatheca containing a sperm mass around half of its volume, and (c) empty spermatheca.

Subsequently, males were weighted and their hind tibia length measured as indicative of their expected fitness (Leather, 1988) (three females from the same age group were also measured). These results were subjected to ANOVA ($P < 0.05$) and, when necessary, means were separated using Duncan's test. Immediately after the measuring, males were dissected in a saline solution to check the appearance of the free lobes of their two-bilobed testes (Aslam, 1961). Testicular deterioration was rated according to their appearance: a rating of 100 was assigned when the four testicular lobes were turgid and sparkling, 66.7 when lobes showed lack of turgidity, 33.3 when the four lobes were shrivelled, and 0 when lobes showed protuberances (fig. 1). Mated females were preserved in 70% ethanol for two weeks to allow sperm in their sclerotized spermathecal capsules to coalesce. Then, successful sperm transmission was rated according to the following scale: a rating of 0 was assigned to spermathecae containing no sperm, 50 to those containing a sperm mass around half of the spermathecal volume, and 100 to those nearly full of sperm (fig. 2). The percentage of apparent successful matings, as well as testicular deterioration and successful sperm transmission, were transformed into probits and the corresponding probit lines fitted (Polo Plus, LeOra Software Inc., Berkeley, CA, USA) to estimate how fast irradiated males lost competitiveness relative to control ones. A chi-square test was used to estimate the goodness-of-fit. Based on these lines, times to 50% and 90% (i) apparent successful mating (ASMT₅₀ and ASMT₉₀), (ii) testicular deterioration (TDT₅₀ and TDT₉₀) and (iii) successful sperm transmission (SSTT₅₀ and SSTT₉₀) were determined.

Reproductive capacity of irradiated males

Fecundity and egg hatching of females mated with untreated or 15-Gy gamma-irradiated males of different ages (4, 10, 16, 22 and 28-day-old) were assessed. Six males per treatment and age were selected and allowed to mate with a 15-day-old female as above. This age was used based on results from the Females Reproductive Biology assay. Subsequently, females were transferred to a standard container. Eggs were collected daily and egg hatching was assessed as before for 15 days. Males were kept in separate standard containers; and, when they reached the age 42 days, the same procedure was repeated for eight untreated and eight 15-Gy irradiated males to assess their re-mating success. Means were compared by ANOVA ($P < 0.05$) and separated using Duncan's test.

Semi-field trials

Two semi-field assays were carried out to study *R. ferrugineus* infestation in palms caused by untreated females

Table 1. Preoviposition time (days), fecundity (number of eggs female⁻¹) and egg hatching (%) during the first 30 days of newly emerged females individually introduced into a Petri dish either alone (virgin females) or with a five-day-old male for the first 30 min (male-exposed females). In all cases, figures shown represent mean \pm SE, and minimum and maximum values in parenthesis.

Female status	Preoviposition time		Fecundity*				Egg hatching	
	<i>n</i>	All females	<i>n</i>	All females	<i>n</i>	Unfertile females	<i>n</i>	Fertile females
Virgin	15	6.33 \pm 0.8 (2–13)	15	48.1 \pm 5.6 (13–84)	15	48.1 \pm 5.6 (13–84)	–	–
Male-exposed	15	5.27 \pm 0.4 (3–8)	15	138.5 \pm 16.8 (43–243)	3	46.7 \pm 2.0 (43–50)	12	161.5 \pm 12.3 (62–243)
ANOVA		<i>t</i> = 0.6041; <i>P</i> = 0.4337		<i>F</i> = 25.41; <i>df</i> = 1, 29; <i>P</i> < 0.0001		<i>F</i> = 0.07; <i>df</i> = 1, 17; <i>P</i> = 0.7973		NA

n, number

* data subjected to the log(*x*) transformation prior to ANOVA.

that had been previously in contact with one of the following four different sorts of males: (i) untreated males (control), (ii) 15-Gy gamma-irradiated males (γ), (iii) *B. bassiana*-challenged males (*Bb*) and (iv) 15-Gy gamma-irradiated and *B. bassiana*-challenged males ($\gamma + Bb$). These assays took place in August and October 2010. Four-year-old potted *P. canariensis* palms obtained from an officially inspected nursery (European Union, 2007) and, therefore, presumed to be free of *R. ferrugineus* were used in these assays. The stipe of these palms was 0.50 m high and 0.45 m wide. These plants were watered twice per week. The *B. bassiana* strain used was isolated from an infected pupa originally collected in a date palm grove in Spain and is deposited in the Spanish Collection of Culture Types with the accession number CECT-20752 (Dembilio *et al.*, 2010b). Suspensions were prepared as described by these authors and kept in 4°C dark storage before use. Two-day-old *R. ferrugineus* males (either irradiated at age 1 or not) were immersed individually for 90 s in a conidial 0.02%-Tween 80 (Tween® 80, synthesis grade, Scharlau Chemie S.A., Sentmenat, Spain) aqueous suspension adjusted to a final concentration of 1.30×10^8 and 1.14×10^8 conidia ml⁻¹ in the first and second trials, respectively. For each treatment (control, γ , *Bb* and $\gamma + Bb$), three and four palms (=replicates) were considered in the first and second trials, respectively. Palms were individually exposed in separate cages to three three-day-old couples of *R. ferrugineus*, either irradiated and/or *B. bassiana*-challenged or not. Before release, for each treatment, males and females were allowed to mate in a plastic box (20 \times 10 \times 8 cm) for 24 h. Ten days after release, when found, adults were removed from the cage. Those recovered from the *B. bassiana* treatments (*Bb* and $\gamma + Bb$) were individually transferred to a standard container where filter paper and apple pieces were replaced once per week for three weeks. In case of death, the weevil was surface sterilized with 1% sodium hypochlorite followed by three rinses with sterile distilled water, placed on a sterile moistened filter paper in an individual sterile Petri dish sealed with Parafilm-M® and kept at room temperature to ascertain the presence of *B. bassiana*. One month after release, palms were carefully dissected and checked for the presence of *R. ferrugineus*. The number of live immature stages per palm was subjected to a 2-way ANOVA. Because of the high relationship observed between temperature and *R. ferrugineus* oviposition, development and survival (Dembilio & Jacas 2011; Dembilio *et al.*, 2012), mean temperature during the assay was considered as a factor. Efficacies (Abbott, 1925) were calculated for those treatments exhibiting significant differences when compared to control.

Results

Laboratory trials

Females reproductive biology

Pre-oviposition time was not significantly different for virgin and mated females (5.5 \pm 0.4 days). Although fecundity showed a great variation among individuals (from 13 to 84 and from 43 to 243 eggs for virgin and mated females, respectively), females exposed to a male laid significantly more eggs than virgin females (138.5 vs. 48.1 eggs per female) (table 1). Likewise, egg hatching of mated females (69.0 \pm 9.9%) and daily oviposition rate (5.6 \pm 0.7 eggs per female and day) were higher than those of unmated females (no egg hatching and 2.0 \pm 0.2 eggs per female and day). On the other hand, three mated females (20% of total females tested) produced no fertile eggs and their fecundity was similar to that of virgin females (table 1). When these three females were excluded from the counts, fecundity of mated females increased to 161.5 eggs, egg hatching to 86.3 \pm 3.5% and the daily oviposition rate to 6.8 \pm 1.0 eggs per female and day (table 1). A sharp increase in the daily rate of oviposition from day 5 to 15 could be observed in mated females (fig. 3). Afterwards, their daily fecundity remained fairly constant, around 9–10 eggs per day, whereas that of unmated females stayed around two eggs. Egg hatching of mated females followed the same trend (fig. 3).

Adult longevity

Untreated males had the longest longevity (244.7 \pm 16.1 days) and were followed by untreated females (197.2 \pm 17.6 days), 15-Gy-treated males (118.6 \pm 13.0 days) and finally 25-Gy-treated males (21.8 \pm 3.6 days) (table 2).

Mating success of irradiated males

Hind tibia length was the same for control insects (6.10 \pm 0.05 mm) irrespective of age (*F* = 1.52; *df* = 15, 92; *P* = 0.1197) and sex (*F* = 0.21; *df* = 1, 92; *P* = 0.6449). Similarly, body weight was independent of age (*F* = 1.54; *df* = 15, 92; *P* = 0.1111) but significantly changed with sex (*F* = 9.30; *df* = 1, 92; *P* = 0.0032) and resulted in 1.14 \pm 0.04 and 1.00 \pm 0.003 g for females and males, respectively. When tibia length and weight of control and irradiated males were compared, no differences were obtained for the 16 ages and the three doses considered (*P* \geq 0.4283 in all cases). However, the higher the dose, the sooner testes lost turgidity (table 3). In this case, the three curves obtained for testicular deterioration (TD) (fig. 4) could

Table 2. Longevity (days; mean \pm SE) of *R. ferrugineus* adults: untreated females and males, and gamma-irradiated males.

Longevity	Females b	Males a	15-Gy irradiated males c	25-Gy irradiated males d
1st assay (max–min)	185.4 \pm 18.4 (59–305)	246.5 \pm 17.6 (103–371)	118.7 \pm 25.6 (20–323)	21.4 \pm 1.9 (11–44)
2nd assay (max–min)	209.0 \pm 19.0 (49–347)	242.9 \pm 21.0 (54–380)	118.7 \pm 25.1 (20–360)	22.3 \pm 2.8 (11–58)
ANOVA F; df; P	Assay # Sex and treatment Interaction	0.16; 1, 135; 0.6908 55.43; 3, 135; <0.0001 0.23; 3, 135; 0.8785		

Column headings followed by the same letter are not significantly different.

Table 3. Probit lines adjusted to different parameters measuring mating success of *R. ferrugineus* couples (ASM, percentage apparent successful mating; TD, percentage testicular deterioration; SST, percentage successful sperm transmission) with different gamma-irradiation treatments (control, 15 Gy and 25 Gy gamma-irradiated males) along time. Time is expressed in weeks.

Treatment	N	Slope	df	X ²	P	xT ₅₀	95% f.l.	xT ₉₀	95% f.l.
Apparent successful mating						ASMT₅₀		ASMT₉₀	
25-Gy	12	3.166 \pm 1.835	2	4.152	0.1254	<3	–	<3	–
15-Gy	36	2.304 \pm 1.087	9	5.267	0.8104	>24	–	>24	–
Control	48	0.553 \pm 0.462	14	13.021	0.5249	>52	–	>52	–
Testicular deterioration						TDT₅₀		TDT₉₀	
25-Gy	12	4.771 \pm 1.577	2	0.434	0.8049	2.709	2.001–3.707	5.028	3.684–16.980
15-Gy	36	5.404 \pm 0.953	10	4.838	0.9017	11.986	10.169–14.361	20.692	16.759–30.122
Control	48	4.674 \pm 0.846	14	6.272	0.9592	24.225	20.438–28.904	45.544	36.450–68.705
Successful sperm transmission						SST₅₀		SST₉₀	
25-Gy	12	4.727 \pm 1.755	2	1.106	0.5752	<2	–	<2	–
15-Gy	36	4.312 \pm 0.902	10	9.759	0.4619	8.098	6.498–10.423	16.052	12.030–29.161
Control	48	–	14	32.165	0.0038	24 > x > 20	–	>52	–

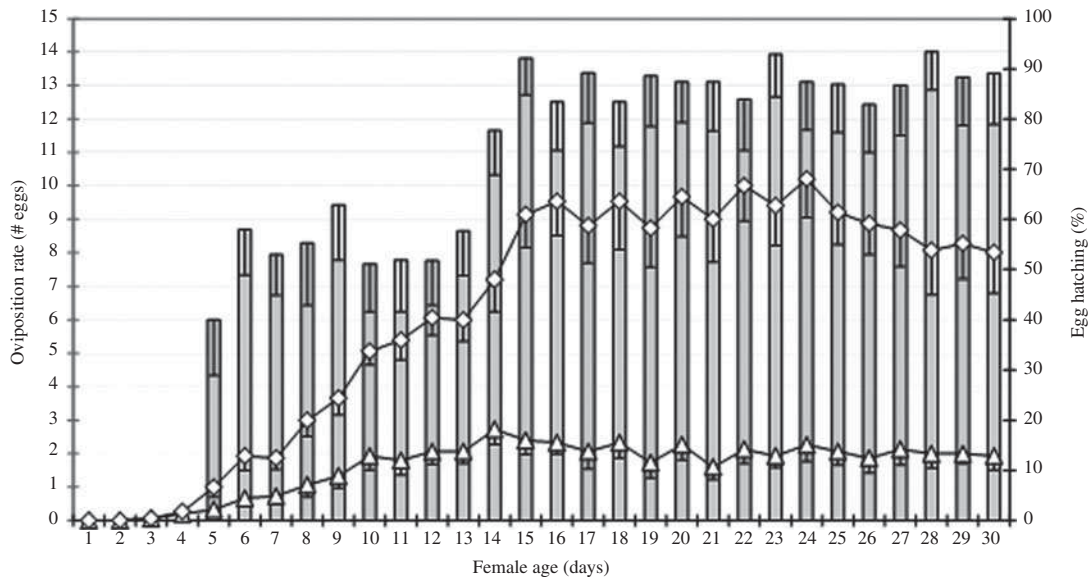


Fig. 3. Daily oviposition rate (eggs per female per day: lines) and percentage egg hatching (eggs per female per day: bars) of male-exposed (grey bars and white squares) and virgin (white triangles) females of *R. ferrugineus*. No egg hatching was registered for virgin females. Only negative error bars are presented to avoid confusion.

be satisfactorily forced to parallelism (chi-square: 0.35; df=2; $P=0.840$) and the relative potency of 15-Gy irradiation was 2.012 and that of 25-Gy, 8.942 (testicular deterioration time 90, TDT₉₀, for control males = 45.5 weeks). In the case of successful sperm transmission (SST), from the three categories established to classify spermathecae (fig. 2), only two, empty and

almost full spermathecae, were actually observed. In any case, SST followed the same trend as TD, (successful sperm transmission time 90, SST₉₀ < 2 weeks for 25-Gy irradiated males and SST₉₀ > 52 weeks for control), probit analysis could be satisfactorily applied to 15-Gy gamma-irradiated males only (table 3). Interestingly, the percentage of apparent

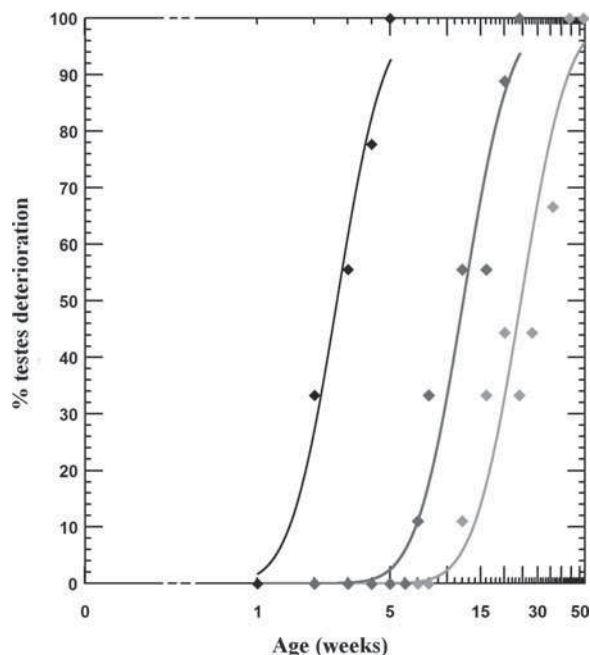


Fig. 4. Probit curves for the deterioration time (in weeks) of testes of control (light grey), 15-Gy (dark grey) and 25-Gy (black) gamma-irradiated males of *R. ferrugineus*.

successful matings sharply decreased along time for 25-Gy irradiated males (no apparent successful mating at age four weeks) but remained almost the same for control and 15-Gy irradiated males until week 24, when the last survivors from the 15-Gy treatment were tested (table 3).

Reproductive capacity of irradiated males

There were no significant differences in the fecundity of females, regardless of the age and the irradiation status of males that they mated with (table 4). However, egg hatching was significantly reduced by male irradiation (87.0 ± 1.7 versus $31.4 \pm 1.7\%$ for control and 15 Gy-gamma irradiated males, respectively). To ascertain whether this reduction was due to the effects of irradiation on sperm quality or on male sexual competitiveness, we further analysed fecundity in fertile and unfertile couples separately. The number of unfertile couples was the same irrespective of the irradiation status of males (30%) and the fecundity of unfertile (29.9 ± 4.6 eggs per female) and fertile couples (86.8 ± 5.5 eggs per female) did not depend on the irradiation status of males. Therefore, possible effects of irradiation on male sexual competitiveness could be discarded. Further, egg hatching did not decrease when control males were allowed to mate with a second female but significantly declined for irradiated males ($85.9 \pm 2.8\%$ vs. $6.1 \pm 0.5\%$ for control and irradiated males, respectively; $F = 806.34$; $df = 1, 15$; $P < 0.0001$). Remarkably, fecundity did not change in this case either ($F = 0.87$; $df = 1, 15$, $P = 0.3660$).

Semi-field trials

Ten days after release, $44.8 \pm 5.5\%$ males and $24.3 \pm 4.0\%$ females could be recovered from infested palms. During the

subsequent three weeks, $51.0 \pm 9.4\%$ males and $38.3 \pm 15.3\%$ females from the fungus-challenged treatments showed post-mortem endogenous hyphal growth of *B. bassiana*.

Both male treatment and temperature affected the number of *R. ferrugineus* immature stages found in *P. canariensis* palms, but the interaction between these two factors was not significant (table 5). The number of *R. ferrugineus* immature stages found was highest when females mated with untreated males (control). Lowest values were obtained for combinations including irradiated males and intermediate values when males had been fungus-challenged only (table 5). According to these values, highest efficacies were obtained for irradiated males, either fungus challenged ($86.5\text{--}100\%$) or not ($88.0\text{--}94.4\%$), which were significantly higher than those observed for fungus-challenged only males ($44.4\text{--}62.6\%$). In this case, temperature did not affect results and efficacies where the same in August and October.

Discussion

Wattanapongsiri (1966) found that *R. ferrugineus* adult females were capable of laying eggs on the day after emergence. Our results prove that a minimum of two days are necessary for females to start oviposition, both virgin and mated females (table 1), and this is in agreement with results from Kaakeh (2005). From that day on, egg laying increased until reaching a roughly constant oviposition rate at age 14 days of about two and 9–10 eggs per female and day for virgin and mated females, respectively. Higher fecundity of mated females is indicative that mating is required to promote egg production in *R. ferrugineus*. Remarkably, the oviposition rate of mated females was much higher than that reported in previous studies using laboratory-reared specimens (2.48 eggs per female and day; Dembilio *et al.*, 2012). The high nutritional value of *P. canariensis* relative to that of meridic diets (Dembilio & Jacas, 2011) could be behind these differences, and this result should be taken into account when extrapolating laboratory results to field conditions. As previously reported (Wattanapongsiri, 1966), fecundity in *R. ferrugineus* females, either mated or not, is characterized by a high variation among individuals (table 1). During the 30 days that our experiment lasted, mated females laid from 43 to 243 eggs with a mean of 138.5. Viado & Bigornia (1949) found that more than 80% of total egg production in *R. ferrugineus* occurred during the first three weeks of life. Therefore, our results should be a rough estimate of total fecundity, and this is in agreement with results from other authors (Butani, 1975; Kranz *et al.*, 1982; Avand-Faghih, 1996). However, higher fecundities, up to 800 eggs per female, have also been reported (Lepesme, 1947; Wattanapongsiri, 1966).

The 30 minute mating time used in our assays, the same as Al-Ayedh & Rasool (2009), resulted in about 80% of total females exposed to control males laying fertile eggs. Egg hatching for these females was higher than 85% in all our assays, and these values are a bit higher than those reported in similar studies (around 75%: Dembilio *et al.*, 2010b; Ll  cer *et al.*, 2010). This percentage, though, was nil for virgin females, and this means that *R. ferrugineus* cannot reproduce parthenogenetically (fig. 3 and table 1). This fact, together with longevity and mating success of sterile males relative to wild insects, are critical points for the success of SIT but also for the purpose of using irradiated males for dissemination of *B. bassiana* infective propagules among wild population.

Table 4. Mean fecundity (mean, number of eggs \pm SE) and percentage egg hatching of 15-day-old *R. ferrugineus* females during the 15 days following copulation with either control or 15-Gy gamma-irradiated males of different ages (days).

Treatment	Male Age	Fecundity*						Egg hatching Fertile females
		<i>n</i>	All females	<i>n</i>	Unfertile females	<i>n</i>	Fertile females	
Control	4	6	72.3 \pm 19.9	2	24.5 \pm 11.5	4	96.3 \pm 20.0	87.8 \pm 2.7
	10	6	76.2 \pm 13.0	2	62.0 \pm 10.0	4	83.3 \pm 18.9	87.4 \pm 5.3
	16	6	66.2 \pm 13.7	2	32.5 \pm 23.5	4	83.0 \pm 9.6	86.9 \pm 3.0
	22	6	39.5 \pm 11.4	2	10.0 \pm 6.0	4	54.3 \pm 10.1	87.4 \pm 4.8
	28	6	57.5 \pm 12.7	1	27.0	5	63.6 \pm 13.6	85.8 \pm 4.3
15-Gy	4	6	47.2 \pm 14.0	4	26.7 \pm 7.3	2	88.0 \pm 10.0	34.6 \pm 10.3
	10	6	102.3 \pm 21.0	1	17.0	5	119.4 \pm 15.1	30.7 \pm 2.7
	16	6	85.7 \pm 13.5	1	21.0	5	98.6 \pm 4.7	34.7 \pm 3.9
	22	6	78.0 \pm 14.0	1	53.0	5	83.0 \pm 16.0	25.8 \pm 2.3
	28	6	72.3 \pm 23.2	2	27.5 \pm 1.5	4	94.8 \pm 29.0	33.7 \pm 2.9
ANOVA (F, df, P)	Irradiation	1.59; 1, 59; 0.2135		0.13; 1, 17; 0.7261		4.00; 1, 41; 0.0541		114.58; 1, 41; <0.0001
	Male Age	1.13; 4, 59; 0.3510		0.19; 4, 17; 0.9386		1.57; 4, 41; 0.2065		0.77; 4, 41; 0.5511
	Interaction	1.22; 4, 59; 0.3135		1.71; 4, 17; 0.2410		0.33; 4, 41; 0.8535		0.81; 4, 41; 0.5296

* data subjected to the log(x) transformation prior to ANOVA.

Table 5. Mean number of live immature stages of *R. ferrugineus* (\pm SE) found in *P. canariensis* and efficacies (%) as a function of the mean monthly temperature (T, °C) during the trial and male treatment: (i) untreated males (control), (ii) gamma-irradiated males (15 Gy), (iii) *B. bassiana*-challenged males (*Bb*), and (iv) gamma-irradiated *B. bassiana*-challenged males (15 Gy + *Bb*). Before release, three males and three untreated females per palm were allowed to mate for a day. Palm dissection took place one month after release.

Assay	T	Male treatment	<i>n</i>	No. immature stages*	Efficacy	
August	25.0±0.3	Control	3	22.3±11.0	–	
		15 Gy	3	2.7±2.2	88.0±9.8	
		<i>Bb</i>	3	8.3±4.8	62.6±21.7	
		15 Gy+ <i>Bb</i>	3	3.0±1.1	86.5±5.2	
October	15.4±0.4	Control	4	4.5±1.0	–	
		15 Gy	4	0.3±0.3	94.4±5.5	
		<i>Bb</i>	4	2.5±0.6	44.4±14.3	
		15 Gy+ <i>Bb</i>	4	0	100	
ANOVA (<i>F</i> , <i>df</i> , <i>P</i>)		Male treatment	13.44; 3, 27; <0.0001		8.04; 2, 20; 0.0042	
		Temperature	20.79; 1, 27; 0.0002		0.01; 1, 20; 0.9519	
		Interaction	0.44; 3, 27; 0.7248		1.11; 2, 20; 0.3557	
Mean separation (Duncan; <i>P</i> < 0.005)				control> <i>Bb</i> > 15 Gy= 15 Gy+ <i>Bb</i> T _{august} > T _{october}		<i>Bb</i> < 15 Gy= 15 Gy+ <i>Bb</i>

* data subjected to the log (x + 1) transformation prior to analysis.

An earlier study by Rahalkar *et al.* (1973) reported that treatment of newly emerged sugarcane-reared *R. ferrugineus* males at a dose of 15 Gy resulted in 90% sterility in males with no adverse effect on survival, whereas higher doses increased sterility at the expense of reducing survival. Because immature nutrition, radiation dose and their interaction significantly affect male lifespan and the hatchability of eggs resulting of mating with wild females (Al-Ayedh & Rasool, 2010), we decided to include in our study 15 Gy as well as 25 Gy, the lowest dose used by these authors in a study with gamma-irradiated males under variable relative humidity (Al-Ayedh & Rasool, 2009). The short longevity (table 2) and fast effect on testes (table 3 and fig. 4) observed at 25 Gy made us discard this dose in subsequent assays. Short longevity might be related to damage in the dividing regenerative cells of the mid-gut, which are particularly sensitive to radiation (Mohamed, 2010). In fact, necrotic segments of intestine were frequently observed during dissection of 25-Gy irradiated males. Likewise, the accelerated deterioration of testes in gamma-irradiated males could be related to injury in testicular tissues.

Al-Waneen *et al.* (2009) observed damaged spermatid tubules, lysed spermatid cysts and the breakage at the junction of spermatid tubes and vas efferens that disconnected the normal passage of sperm in irradiated *R. ferrugineus* males. Although 15-Gy irradiated males had their testes damaged and, as a consequence, successful sperm transmission decreased faster than in control males (table 3), the percentage of apparent successful matings remained almost the same as for control males during the 24 weeks that 15-Gy irradiated male survivors could be tested. The longevity of 15-Gy irradiated males was considerably longer than that reported by other authors using different food sources (Rahalkar *et al.*, 1973; Prabhu & Patil, 2009; Al-Ayedh & Rasool, 2010) and, once again, this may be related to the high nutritional value of *P. canariensis* relative to other food sources (Dembilio & Jacas, 2010). Although 15-Gy irradiated males lived about half the time that untreated males did (120 vs. 245 days; table 2), this lifespan would be enough to ensure that irradiated males, if infected with EPF, had the time enough to transmit the infective fungus propagules to wild females and their

descendants, as well as to disperse and act themselves as an inoculum source after death (Dembilio *et al.*, 2010b). This hypothesis could be proved later during the semi-field trials (table 5).

Fecundity of fertile females mated with 15-Gy gamma-irradiated males did not change relative to control (86.8 eggs; table 4). However, egg hatching in couples where males had been irradiated was lower than in control (31.4% vs. 87.0%; table 4), and this value decreased even more after a second mating (6.1% vs. 85.9%). The higher egg viability for irradiated males on the first mating was predictable because mature spermatozoa present at the time of irradiation are radio-resistant (Rahalkar *et al.*, 1975). In any case, this reduction undoubtedly contributed to the results obtained in the semi-field trials where infestation in control palms was always the highest (table 5). However, infestation significantly changed during the two semi-field assays included in our study. The number of immature stages found during the first assay (August 2010) was similar to that found in an assay carried out in summer 2009 using the same infestation method (22.8 ± 5.0 specimens; mean temperature 23.7  C) (Ll  cer *et al.*, 2010). Temperature affects oviposition, development and immature survival of *R. ferrugineus* (Dembilio & Jacas, 2011; Dembilio *et al.*, 2012), and therefore it could explain the lower number of immature stages found in October relative to August. Anyway, the significance of the differences observed among treatments was the same in both semi-field trials (table 5). Highest efficacies were obtained for treatments with irradiated males, irrespective of whether the males had been fungus-challenged or not. However, even in these cases, infestation occurred. This result was expected because, as mentioned before, egg hatching in combinations where males had been irradiated was not nil. Furthermore, the larvae obtained from these couples are not expected to suffer any differential mortality from control ones. However, those obtained from fungus-challenged treatments are (Dembilio *et al.*, 2010b). Unfortunately, the destructive sampling used in our assays did not allow us to keep the immature stages found in the palms alive to check whether they were infected by *B. bassiana*. Nevertheless, we could see that almost 40% of the females recovered from fungus-challenged couples were actually infected and died from this infection. This horizontal transmission had already been reported by Dembilio *et al.* (2010b), and these authors also reported lethal effects on the offspring. Therefore, our assays probably underestimated the efficacy of the combinations where *B. bassiana* was involved (*Bb* and γ +*Bb*). A continuous monitoring of the insects emerging from the palms could allow a proper estimation of this effect. This result should be taken into account in future experiments aimed at measuring the efficacy of this type of treatments.

Our results suggest that *R. ferrugineus* infestation in palms could be reduced by the release of gamma-irradiated *B. bassiana*-challenged males in the field. This reduction would be caused by the joint effect of the low offspring production of females mated with irradiated males and the transmission of *B. bassiana* from fungus-challenged males. Therefore, the inoculation of *B. bassiana* on gamma irradiated males could considerably improve the results obtained in the field using sterilized males only (Rahalkar *et al.* 1977; Krishnakumar & Maheswari, 2007) thanks to the added effect of the EPF. Because *R. ferrugineus* is polygamous and highly gregarious (Wattanapongsiri, 1966), the combined effects of irradiation and *B. bassiana* infection could be obtained in numerous

specimens from a relative low number of treated males. Fungal application by releasing infected males has been already attempted against other pest insects, among them coleopterans such as sap and bark beetles (Dowd & Vega 2003; Kreutz *et al.*, 2004), with successful results. This auto-dissemination methodology was even tested in the field against *R. ferrugineus*. The number of trap-captured adults decreased when the release of fungus-infected males was combined with the application of *B. bassiana* conidiospores by dusting on date palms (Sewify *et al.*, 2009). This result is especially important if we keep in mind that adult *R. ferrugineus* often remain and extensively reproduce within the same host. However, this behaviour that could hamper the efficacy of this method does not seem to actually do it.

In summary, our results show that the release of irradiated males infected with a suitable strain of *B. bassiana* should be considered as a practical way to improve the biological control of *R. ferrugineus* with EPF.

Acknowledgments

The authors thank J.R. Faleiro (Goa, India) for critical review of the manuscript. Special thanks are extended to E. Quesada-Moraga (University of Cordoba) for the suspensions of *B. bassiana* strain, R. Argil  s (TRAGSA) for the gamma radiation application, and the palm nurserymen association ASFPLANT for providing the palms used in our assays. This research was partially funded by the Spanish Ministerio de Ciencia e Innovaci  n (MICINN project TRT2006-00016-C07-05) and the Valencian Conselleria d'Agricultura, Pesca i Alimentaci   (IVIA project 5611). E. Ll  cer was recipient of a postdoctoral fellowship from the MCINN (Juan de la Cierva program, co-funded by the European Social Fund).

References

- Abbas, M.S.T., Hanounik, S.B., Mousa, S.A. & Mansour, M.I. (2001a) On the pathogenicity of *Steinernema abbasi* and *Heterorhabditis indicus* isolated from adult *Rhynchophorus ferrugineus* (Coleoptera). *International Journal of Nematology* **11**, 69–72.
- Abbas, M.S.T., Saleh, M.M.E. & Akil, A.M. (2001b) Laboratory and field evaluation of the pathogenicity of entomopathogenic nematodes to the red palm weevil, *Rhynchophorus ferrugineus* (Oliv.) (Col.: Curculionidae). *Journal of Pest Science* **74**, 167–168.
- Abbas, M.S.T., Hanounik, S.B., Shahdad, A.S. & Al-Bagham, S.A. (2006) Aggregation pheromone traps, a major component of IPM strategy for the red palm weevil, *Rhynchophorus ferrugineus* in date palms (Coleoptera: Curculionidae). *Journal of Pest Science* **79**, 69–73.
- Abbott, W.S. (1925) A method of computing the effectiveness of an insecticide. *Journal of Economical Entomology* **18**, 265–267.
- Al-Ayedh, H.Y. & Rasool, K.G. (2009) Sex ratio and the role of mild relative humidity in mating behaviour of red date palm weevil *Rhynchophorus ferrugineus* Oliv. (Coleoptera: Curculionidae) gamma-irradiated adults. *Journal of Applied Entomology* **134**, 157–162.
- Al-Ayedh, H.Y. & Rasool, K.G. (2010) Determination of the optimum sterilizing radiation dose for control of the red date palm weevil *Rhynchophorus ferrugineus* Oliv. (Coleoptera: Curculionidae). *Crop Protection* **29**, 1377–1380.
- Al-Waneen, W.S., Al-Kalifah, M.S. & Al-Ayedh, H.Y. (2009) Effect of gamma radiation on the histology of the testis of red

- date palm weevils, *Rhynchophorus ferrugineus* (Olivier). *Arab Journal of Plant Protection* **27**, 20.
- Aslam, N.A.** (1961) An assessment of some internal characters in the higher classification of the Curculionidae s.l. (Coleoptera). *Transactions of the Royal Entomological Society of London* **113**, 417–480.
- Avand-Faghih, A.** (1996) The biology of red palm weevil, *Rhynchophorus ferrugineus* Oliv. (Coleoptera: Curculionidae) in Saravan region (Sistan and Balouchistan Province, Iran). *Applied Entomology and Phytopathology* **63**, 16–18.
- Butani, D.K.** (1975) Insect pests of fruit crops and their control. 15: Date palm. *Pesticides* **9**, 40–42.
- Dembilio, Ó. & Jacas, J.A.** (2011) Basic bio-ecological parameters of the invasive Red Palm Weevil, *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae), in *Phoenix canariensis* under Mediterranean climate. *Bulletin of Entomological Research* **101**, 153–163.
- Dembilio, Ó., Jacas, J.A. & Llácer, E.** (2009) Are the palms *Washingtonia filifera* and *Chamaerops humilis* suitable hosts for *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae)? *Journal of Applied Entomology* **133**, 565–567.
- Dembilio, Ó., Llácer, E., Martínez de Altube, M.M. & Jacas, J.A.** (2010a) Field efficacy of Imidacloprid and *Steinernema carpocapsae* in a chitosan formulation against the Red Palm Weevil *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae) in *Phoenix canariensis*. *Pest Management Science* **66**, 365–370.
- Dembilio, Ó., Quesada-Moraga, E., Santiago-Álvarez, C. & Jacas, J.A.** (2010b) Potential of an indigenous strain of the entomopathogenic fungus *Beauveria bassiana* as a biological control agent against the Red Palm Weevil, *Rhynchophorus ferrugineus*. *Journal of Invertebrate Pathology* **104**, 214–221.
- Dembilio, Ó., Tapia, G., Téllez, M.M. & Jacas, J.A.** (2012) Lower temperature thresholds for oviposition and egg hatching of the Red Palm Weevil, *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae), in a Mediterranean climate. *Bulletin of Entomological Research* **102**, 97–102.
- Dowd, P.F. & Vega, F.E.** (2003) Autodissemination of *Beauveria bassiana* by Sap Beetles (Coleoptera: Nitidulidae) to overwintering sites. *Biocontrol Science Technology* **13**, 65–75.
- Elawad, S.A., Mousa, S.A., Shahdad, A.S., Alawaash, S.A. & Alamiri, A.M.A.** (2007) Efficacy of entomopathogenic nematodes against red palm weevil in UAE. *Acta Horticulturae* **736**, 415–420.
- El-Sufy, R., Al-Awash, S.A., Al-Bgham, S., Shahdad, A.S. & Al-Bathra, A.H.** (2009) Pathogenicity of the fungus *Beauveria bassiana* (Bals.) Vuill to the Red Palm Weevil, *Rhynchophorus ferrugineus* (Oliv.) (Col.: Curculionidae) under Laboratory and Field Conditions. *Egyptian Journal of Biological Pest Control* **19**, 81–85.
- European and Mediterranean Plant Protection Organization** (2009) First record of *Rhynchophorus ferrugineus* in Curaçao, Netherlands Antilles. *European and Mediterranean Plant Protection Organization Reporting Service*. Pests & Diseases. 2009/002. Available online at <http://archives.eppo.int/EPPORreporting/2009/Rsf-0901.pdf>.
- European Union** (2007) Commission Decision 2007/365/EC of 25 May 2007 on emergency measures to prevent the introduction into and the spread within the Community of *Rhynchophorus ferrugineus* (Olivier). *Official Journal European Union* **L139**, 24–27.
- Faleiro, J.R.** (2006) A review of the issues and management of the red palm weevil *Rhynchophorus ferrugineus* (Coleoptera: Rhynchophoridae) in coconut and date palm during the last one hundred years. *International Journal of Tropical Insect Science* **26**, 135–154.
- Faleiro, J.R. & Kumar, J.A.** (2008) A rapid decision sampling plan for implementing area-wide management of the red palm weevil, *Rhynchophorus ferrugineus*, in coconut plantations of India. *Journal Insect Science* **8**. Article 15. Available online at <http://insectscience.org/8.15> (accessed 21 August 2012)
- Gindin, G., Levski, S., Glazer, I. & Soroker, V.** (2006) Evaluation of the entomopathogenic fungi *Metarhizium anisopliae* and *Beauveria bassiana* against the Red Palm Weevil *Rhynchophorus ferrugineus*. *Phytoparasitica* **34**, 370–379.
- Gothi, K.K., Hire, R.S., Vijayalakshimi, N. & Dongre, T.K.** (2007) Studies on mating behaviour of radio-sterilized males of red palm weevil, *Rhynchophorus ferrugineus* (Oliv). *Journal of Nuclear Agriculture and Biology* **36**, 65–72.
- Kaakeh, W.** (2005) Longevity, fecundity, and fertility of the red palm weevil, *Rhynchophorus ferrugineus* Olivier (Coleoptera: Curculionidae) on natural and artificial diets. *Emirates Journal of Agricultural Sciences* **17**, 23–33.
- Kranz, J.H., Schmutterer, H. & Koc, W.** (1982) *Enfermedades, Plagas y Malezas de los Cultivos Subtropicales*. Berlin, Germany, Verlag Paul Parey.
- Krishnakumar, R. & Maheswari, P.** (2007) Assessment of the Sterile Insect Technique to Manage Red Palm Weevil *Rhynchophorus ferrugineus* in Coconut. pp. 475–485 in Vreysen, M.J.B., Robinson, A.S. & Hendrichs, J. (Eds) *Area-wide Control of Insect Pests*. Dordrecht, The Netherlands, Springer.
- Kreutz, J., Zimmermann, G. & Vaupel, O.** (2004) Horizontal Transmission of the entomopathogenic fungus *Beauveria bassiana* among the Spruce Bark Beetle, *Ips typographus* (Col., Scolytidae) in the laboratory and under field conditions. *Biocontrol Science and Technology* **14**, 837–848.
- Lacey, L.A., Kirk, A.A., Millar, L., Mereadier, G. & Vidal, C.** (1999) Ovicidal and larvicidal activity of conidia and blastospores of *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes) against *Bemisia argentifolii* (Homoptera: Aleyrodidae) with a description of a bioassay system allowing prolonged survival of control insects. *Biocontrol Science and Technology* **9**, 9–18.
- Lance, D.R. & McInnis, D.O.** (2005) Biological Basis of the Sterile Insect Technique. pp. 69–94 in Dyck, V.A., Hendrichs, J. & Robinson, A.S. (Eds) *Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management*. Dordrecht, The Netherlands, Springer.
- Leather, S.R.** (1988) Size, reproductive potential and fecundity in insects: things aren't as simple as they seem. *Oikos* **51**, 386–388.
- Lepesme, P.** (1947) *Les Insectes des Palmiers*. Paris, France, P. Lechevalier.
- Llácer, E. & Jacas, J.A.** (2010) Efficacy of phosphine as a fumigant against *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae) in palms. *Spanish Journal of Agricultural Research* **8**, 775–779.
- Llácer, E., Martínez de Altube, M.M. & Jacas, J.A.** (2009) Evaluation of the efficacy of *Steinernema carpocapsae* in a chitosan formulation against the Red Palm Weevil, *Rhynchophorus ferrugineus* in *Phoenix canariensis*. *BioControl* **54**, 559–565.
- Llácer, E., Dembilio, O. & Jacas, J.A.** (2010) Evaluation of the Efficacy of an Insecticidal Paint Based on Chlorpyrifos and Pyriproxyfen in a Micro-encapsulated Formulation against the Red Palm Weevil, *Rhynchophorus ferrugineus*. *Journal of Economical Entomology* **103**, 402–408.

- Llácer, E., Negre, M. & Jacas, J.A. (2012) Evaluation of an oil dispersion formulation of Imidacloprid as a drench against *Rhynchophorus ferrugineus* (Coleoptera, Curculionidae) in young palm trees. *Pest Management Sciences* **68**, 878–882.
- Martín, M.M. & Cabello, T. (2006) Manejo de la cría del picudo rojo de la palmera, *Rhynchophorus ferrugineus* (Olivier, 1790) (Coleoptera, Dryophthoridae), en dieta artificial y efectos en su biometría y biología. *Boletín de Sanidad Vegetal. Plagas* **32**, 631–641.
- Mohamed, H.F. (2010) Histological Studies On The Midgut Of The Gamma Irradiated Red Palm Weevil, *Rhynchophorus ferrugineus* (Olivier) Coleoptera: Curculionidae. *Isotope & Radiation Research* **42**, 161–172.
- Murphy, S.T. & Briscoe, B.R. (1999) The red palm weevil as an alien invasive: biology and the prospects for biological control as a component of IPM. *Biocontrol News and Information* **20**, 35–46.
- Novelo-Rincón, L.F., Montoya, P., Hernández-Ortiz, V., Liedo, P. & Toledo, J. (2009) Mating performance of sterile Mexican fruit fly *Anastrepha ludens* (Dipt., Tephritidae) males used as vectors of *Beauveria bassiana* (Bals.) Vuill. *Journal of Applied Entomology* **133**, 702–710.
- Prabhu, S.T. & Patil, R.S. (2009) Studies on the biological aspects of red palm weevil *Rhynchophorus ferrugineus* (Oliv.). *Karnataka Journal of Agricultural Sciences* **22**, 732–733.
- Prabhu, S.T., Dongre, T.K. & Patil, R.S. (2010) Effect of irradiation on the biological activities of red palm weevil, *Rhynchophorus ferrugineus* Olivier. *Karnataka Journal of Agricultural Sciences* **23**, 186–188.
- Quesada-Moraga, E., Santos-Quiros, R., Valverde-Garcia, P. & Santiago-Alvarez, C. (2004) Virulence, horizontal transmission, and sublethal reproductive effects of *Metarhizium anisopliae* (anamorphic fungi) on the German cockroach (Blattodea: Blattellidae). *Journal of Invertebrate Pathology* **87**, 51–58.
- Rahalkar, G.W., Harwalkar, M.R., Rananavare, H.D., Shantaram, K. & Ayengar, A.R.G. (1973) Laboratory studies on radiation sterilization of *Rhynchophorus ferrugineus* males. *Journal of Plantation Crops* **1**, 141–145.
- Rahalkar, G.W., Harwalkar, M.R. & Rananavare, H.D. (1975) Laboratory studies on sterilization of male *Rhynchophorus ferrugineus*. pp. 261–267 in *Sterility Principle for Insect Control* 1974. International Atomic Energy Agency (Ed.), Vienna, Austria.
- Rahalkar, G.W., Harwalkar, M.R., Rananavare, H.D., Kurgan, C., Abraham, V.A. & Koya, K.M.A. (1977) Preliminary field studies on the control of the red palm weevil, *Rhynchophorus ferrugineus* using radio sterilized males. *Journal of Nuclear Agriculture and Biology* **6**, 65–68.
- Ranavara, H.D., Harwalkar, M.R. & Rahalkar, G.W. (1975) Studies on the feasibility of using sterile females along with sterile males in the control of red palm weevil, *Rhynchophorus ferrugineus* Oliv. (Coleoptera: Curculionidae). pp. 861–868 in *Use of Radiations and Radioisotopes in Studies of Plant Productivity: Proceedings of a Symposium held at G.B. Pant University of Agriculture and Technology, Pantnagar, India*, 12–14 April, 1974, Department of Atomic Energy, Bombay, India.
- Ramachandram, C.P. (1991) Effect of gamma radiation on various stages of red date palm weevil, *Rhynchophorus ferrugineus* Oliv. *Journal of Nuclear Agriculture and Biology* **3**, 218–221.
- Saleh, M.M.E. & Alheji, M. (2003) Biological control of red palm weevil with entomopathogenic nematodes in the eastern province of Saudi Arabia. *Egyptian Journal of Biological Pest Control* **13**, 55–59.
- Sewify, G.H., Belal, M.H. & Al-Awash, S.A. (2009) Use of the entomopathogenic fungus, *Beauveria bassiana* for the biological control of the red palm weevil, *Rhynchophorus ferrugineus* Olivier. *Egyptian Journal of Biological Pest Control* **19**, 157–163.
- Viado, G.B. & Bigornia, A.E. (1949) A biological study of the asiatic palm weevil, *Rhynchophorus ferrugineus* (Olivier) (Curculionidae, Coleoptera). *The Philippine Agriculturist* **33**, 1–27.
- Vickers, R.A., Furlong, M.J., White, A. & Pell, J.K. (2004) Initiation of fungal epizootics in diamondback moth populations within a large field cage: proof of concept for auto-dissemination. *Entomologia Experimentalis et Applicata* **111**, 7–17.
- Wattanapongsiri, A. (1966) A Revision of the Genera *Rhynchophorus* and *Dynamis* (Coleoptera: Curculionidae). Bangkok, Thailand. *Department of Agriculture Sciences Bulletin* **1**, 1–329.